**TapeStation D1000 DNA ScreenTape Assay**

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1. Allow TapeStation reagents (stored at 4 ºC) to come to room temperature for 30 min.
2. Turn on the TapeStation and launch the Controller software.
3. Open a new D1000 ScreenTape (or use one with lanes remaining) and flick it gently to remove any air bubbles. ScreenTapes are stored at 4 ºC.
4. Insert the ScreenTape into the ScreenTape nest on the instrument, with the barcode facing back on the right-hand side. The software will recognize the tape and the number of available lanes.
5. Select the required number of sample positions in the software and add sample IDs. The software will display the required number of consumables (tips, tube strips, tapes).
6. Vortex and spin down reagents.
7. Prepare ladder: for 1-2 tapes, add 3 µL D1000 Sample Buffer and 1 µL D1000 Ladder at position A1 in a tube strip. For >2 tapes, add 6 µL buffer and 2 µL ladder to position A1.
8. Add 3 µL of buffer to the remaining tubes for each sample, then add 1 µL of DNA sample to the respective tube.
9. Cap the tube strip(s), then vortex on the IKA MS3 vortexer at 2000 rpm for 1 min (just load and press start). Quick spin to bring everything to the bottom.
10. Load tube strips into the TapeStation, with the ladder at position A1.
11. Remove the strip caps carefully and check that the liquid is still at the bottom.
12. Select **Start**
13. Once complete, the Analysis software will open, and a project file is automatically saved.
14. Empty the tip waste bucket and remove the tape. If the tape still has remaining lanes, put it back in 4 ºC with the reagents for up to 2 weeks. Return all reagents to 4 ºC.

**TapeStation Consumable Prep:**

TapeStation tips (insert new rack when empty)

TapeStation strip tubes and caps (enough for samples +1 ladder)